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# Chiral ligands derived from abrine. Part 5: Substituent effects on asymmetric induction in enantioselective addition of diethylzinc to benzaldehyde catalyzed by chiral oxazolidines possessing an indole moiety

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#### Abstract

A number of the indole-containing chiral oxazolidines possessing the gem-di-p-tolyl and gem-di-o-tolyl groups at  $C_5$  were synthesized from abrine and the effects of the  $C_5$  and  $C_2$  substituents on the asymmetric induction in catalytic enantioselective addition of diethylzinc to benzaldehyde were examined. A working model is proposed to rationalize the asymmetric catalysis by these chiral oxazolidines. © 1998 Elsevier Science Ltd. All rights reserved.

#### 1. Introduction

Enantioselective addition of dialkylzinc to aldehydes catalyzed by chiral β-amino alcohols¹ and other chiral promoters is one of the most extensively investigated asymmetric C-C bond formation reactions in recent years. Extension of this catalytic enantioselective reaction to ketones² and C=N³ functionalities further widens its application to the synthesis of chiral alcohols possessing a stereogenic quaternary carbon center and chiral amines. Our recent work on the alkaloid-based asymmetric synthesis has produced several novel classes of chiral promoters containing an indole moiety.⁴ We found that the chiral oxazolidines 1a-j and related compounds, although lacking a hydroxyl group, could catalyze the reaction of Et<sub>2</sub>Zn with benzaldehyde in up to 53.8% ee.⁴b We describe herein the synthesis and catalysis of chiral oxazolidines 2a-g having the *gem*-di-o-tolyl groups at C<sub>5</sub> and propose a working model to discuss the substituent effect on the catalysis of chiral oxazolidines 1a-j and 2a-g (Fig. 1).

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Fig. 1.

#### 2. Results and discussion

Chiral oxazolidines 1<sup>4b</sup> and 2 were synthesized from the alkaloid, abrine [(S)-N-methyltryptophan]<sup>5</sup> isolated from the seeds of *Abrus precatorius* collected in the Yunnan Province of China (Scheme 1). Reaction of the methyl ester of abrine with excess *p*- or *o*-TolMgBr afforded the corresponding tertiary β-amino alcohols 3a and 3b<sup>4a</sup> which were condensed with a number of aldehydes in the presence of 4 Å MS under very mild conditions (CH<sub>2</sub>Cl<sub>2</sub>, rt or PhH, refluxing) to provide chiral *cis*-2,4-disubstituted oxazolidines 1a-j and 2a-g in 59-89% yield (Table 1). The *trans*-isomers of the oxazolidines were not detected from the condensation reaction in most cases.

Table 1
Synthesis of chiral oxazolidines 1 and 2 from amino alcohols 3a and 3b

Entry	R	1 (Ar = <i>p</i> -Tol)	from 3a	2 (Ar = <i>o</i> -Tol) from <b>3b</b>		
		Method; t; Yield(%)a	[α] <sub>D</sub> <sup>20</sup> (c) <sup>b</sup>	Method; t; Yield(%)a	$[\alpha]_D^{22} (c)^b$	
а	Me	A; 50 h; 88	-12.4 (1.37)	B; 14 h; 59	-134.2 (0.18)	
b	Et	B; 24 h; 67	-60.5 (1.24)	B; 14 h; 76	-199.0 (1.24)	
С	<i>n</i> -Pr	A; 35 h; 85	-56.0 (1.71)	B; 14 h; 77	-207.1 (0.73)	
d	<i>n</i> -Bu	A; 48 h; 65	-50.3 (1.89)	B; 14 h; 76	-207.6 (1.89)	
e	<i>n</i> -Pent	B; 14 h; 70	-50.0 (0.49) <sup>c</sup>	B; 14 h; 71	-206.1 (0.41)	
f	<i>i</i> -Pr	A; 48 h; 62	-105.5 (1.33)	B; 14 h; 85	-205.2 (0.63)	
g	CH <sub>2</sub> - <i>i</i> -Pr	B; 40 h; 89	-67.4 (1.01)	B; 14 h; 70	-219.7 (1.78)	
ħ	CH <sub>2</sub> -t-Bu	B; 60 h; 67	-71.7 (1.08)			
i	(CH <sub>2</sub> ) <sub>2</sub> Ph	A; 40 h; 86	-82.4 (1.58)			
<u>j</u>	Ph	A; 66 h; 79	-96.0 (1.36)			

<sup>&</sup>lt;sup>a</sup>Method A: Reaction was carried out in reflexing PhH; Method B: Reaction was performed in CH<sub>2</sub>Cl<sub>2</sub> at rt. Yields refer to the isolated homogenous materials. <sup>b</sup>Recorded in CHCl<sub>3</sub>. <sup>c</sup>Recorded at 22 °C.

 $Table\ 2$  Reaction of Et<sub>2</sub>Zn with benzaldehyde catalyzed by chiral oxazolidines 1 and 2

Entry	R	Cat*: 1 (Ar = p-Tol) <sup>a</sup>			Cat*: 2 (Ar = o-Tol)		
Entry		Yield(%) <sup>b</sup>	ee(%) <sup>c,e</sup>	Configuration <sup>d</sup>	Yield(%) <sup>b</sup>	ee(%) <sup>c</sup>	Configuration <sup>d</sup>
а	Me	57.3	23.4	S	60.2	5.5	R
b	Et	51.0	26.4	S	34.0	17.2	R
С	n-Pr	57.8	53.8	S	46.0	22.6	R
d	<i>n</i> -Bu	47.2	6.9	S	62.2	8.5	R
е	<i>n</i> -Pent	34.2	3.1	S	69.0	18.2	R
f	<i>i</i> -Pr	56.3	7.4	R	56.7	1.4	R
9	CH <sub>2</sub> - <i>i</i> -Pr	52.7	32.1	s	38.0	4.7	R
h	CH <sub>2</sub> -t-Bu	50.6	28.2	s			
i	(CH <sub>2</sub> ) <sub>2</sub> Ph	52.1	5.3	s			
j	Ph <sup>f</sup>	45.0	0				

<sup>a</sup>Data taken from ref. 4b. <sup>b</sup>Yield is based on the isolated product. Benzyl alcohol was formed in most of the reactions. <sup>c</sup>Determined using HPLC on a CHIRALCEL OD column (FPrOH:Hexane = 95:5, flow rate = 1 mL/min, UV detector at 254 nm). <sup>d</sup>The specific rotation  $[\alpha]_D$  +45.6° (CHCl<sub>3</sub>)<sup>6</sup> for R enantiomer of 4 was used to determine the configuration. <sup>e</sup>The values were reexamined. <sup>f</sup>Reaction for 6 days.

The catalytic potency of 1a-j and 2a-g was examined using the prototype reaction between Et<sub>2</sub>Zn and benzaldehyde with 10 mol% of the catalyst in toluene at room temperature and the results are summarized in Table 2. It is interesting to note that the catalysts 1 induced the formation of the (S)enantiomer of 1-phenyl-1-propanol (-)-4 in up to 53.8% ee and in 34.2-57.8% yield except for chiral oxazolidine 1f bearing an isopropyl group at the  $C_2$  position. In contrast, only the (R)-enantiomer of 1-phenyl-1-propanol (+)-46 was formed in up to 22.6% ee and in 34-69% yield under the catalysis of chiral oxazolidines 2a-g. The opposite asymmetric induction by 1a-e,g-j and 2a-g is influenced by the gem-diaryl groups at C<sub>5</sub>. The enantioselectivity is generally high for C<sub>5</sub> gem-di-p-tolyl-substituted oxazolidines 1 compared with the C<sub>5</sub> gem-di-o-tolyl-substituted analogs 2. A plot of the percentage ee versus the carbon number of the substituent R at C<sub>2</sub> of 1a-e and 2a-e is given in Fig. 2. It is concluded that: (a) the enantiomeric excess varies with the carbon number of R at C<sub>2</sub> with a maximum value recorded for R=n-Pr for both classes of chiral oxazolidines; and (b) the enantioselectivity of the reaction is much more sensitive to the C<sub>2</sub> R group for the catalysts 1a-e compared with that of 2a-e, indicating that the C<sub>2</sub> R group in **1a**-e is in close proximity to the reacting centers in the ethyl-transferring transition state. In other words, a loose transition state operates in the reactions catalyzed by chiral oxazolidines 2a-e and low asymmetric induction is achieved.

Generally speaking, the enantioselectivity induced by chiral oxazolidines 1 and 2 is not very high compared with a variety of hydroxyl group bearing chiral promoters. However, the current work is interesting in the mechanistic aspects of the catalysis. It was reported that chiral diamines 7-9 including

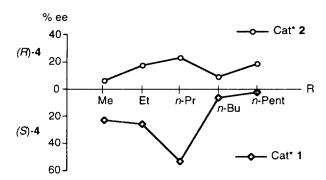


Fig. 2. Relationship between the ees of chiral 1-phenyl-1-propanol (4) and the carbon number of R at the  $C_2$  position of chiral oxazolidine promotors 1 and 2 used for enantioselective ethylation of benzaldehyde

tertiary diamines catalyze the addition reactions of Et<sub>2</sub>Zn to aldehydes by formation of either the N-Zn complexes or zinc amides. 8b.c10.11 The methoxy group in o-anisaldehyde was also reported to form undesirable catalytic species by coordination with Et<sub>2</sub>Zn. <sup>9a</sup> The chiral oxazolidines **1a-j** and **2a-g** have two basic sites, i.e. the nitrogen and oxygen atoms of the oxazolidine ring. If the indole nitrogen could be deprotonated by Et<sub>2</sub>Zn, a third basic site might be available for complexation. Nevertheless, we consider that the nitrogen and oxygen atoms of the oxazolidines contribute primarily to the catalysis through the transition state (TS) 5 (Fig. 3). The oxygen-bound zinc species [(Et<sub>b</sub>)<sub>2</sub>Zn<sub>b</sub>] is much more reactive<sup>9a</sup> and will transfer the Eth group onto the C=O of benzaldehyde complexed with Zna, the latter is also coordinated with the nitrogen atom. An alternative to TS 5 is considered by sharing one Eta group on Zn<sub>a</sub> with Zn<sub>b</sub> to form a polycyclic assembly (a bridged 5/6/6-ring system) which is likely to be too rigid to work. TS 5 predicts the si-face attack at benzaldehyde to give (S)-4 as the major enantiomer which is consistent with our experimental results of 1a-e. 4b Modification on the gem-diaryl groups at C<sub>5</sub> can significantly affect the complexation ability of the oxygen atom. With two bulky o-tolyl groups at C<sub>5</sub>, TS 5 fails to operate because the oxygen atom is incapable of forming a complex due to the severe steric repulsion. Under this circumstance, the Et-transferring zinc species should attack intermolecularly by another zinc complex of the oxazolidine nitrogen or a zinc amide of the indole<sup>8b,c,10,12</sup> at the Zn<sub>a</sub>coordinated benzaldehyde from the re-face (TS 6) to give (R)-4. Due to the great separation among the reacting and the C2/C4 stereogenic centers, the asymmetric induction of 2a-g should be low and less sensitive to the R group at  $C_2$ .

In summary, we have examined a number of chiral oxazolidines 1a–j and 2a–g possessing an indole moiety as promoters in the enantioselective addition of  $Et_2Zn$  to benzaldehyde and found that substituents on both  $C_2$  and  $C_5$  positions of the oxazolidine ring significantly influence the asymmetric induction. Based on these results, we propose a transition state f for catalysis by f0 and f1 featuring the coordination of both oxygen and nitrogen atoms of the oxazolidine with the zinc species. Our finding will encourage further study on the use of readily available chiral oxazolidines f1 in asymmetric catalysis.

# 3. Experimental section

<sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on a Bruker ARX 300 or AM-400 NMR instrument. IR spectra were taken on a Bio-Rad FTS-IR spectrophotometer. Mass spectra (MS) were measured on a Finnigan TSQ 7000 mass spectrometer. High resolution mass spectra (HRMS) were measured by a VG Autospec mass spectrometer under FAB<sup>+</sup> conditions. Elemental analysis was performed on a Model 1106 instrument. Optical rotations were recorded on a Perkin–Elmer 241 polarimeter. All reactions were

Fig. 3.

carried out under a nitrogen atmosphere and monitored by thin-layer chromatography on 0.25 mm E. Merck silica gel plates (60 F-254) using UV light, or 7% ethanolic phosphomolybdic acid and heating as the visualizing methods. E. Merck silica gel 60 (particle size 0.040–0.063 mm) was used for flash column chromatography. Yields refer to chromatographically and spectroscopically (<sup>1</sup>H NMR) homogeneous materials. Abrine was isolated from the extract of the seeds of *Abrus precatorius* collected in the Yunnan Province of China.<sup>5</sup> Amino alcohols **3a** and **3b** were synthesized from abrine according to the known procedure.<sup>4a</sup> Et<sub>2</sub>Zn (1.0 M in hexanes) and other reagents were obtained commercially and used as received.

# 3.1. Oxazolidines **1a-j** and **2a-g**; general procedure

Method A: A solution of **3a** (128 mg, 0.33 mmol) and the aldehyde (0.50 mmol) in dry PhH (10 mL) in the presence of powdered 4 Å MS was heated at refluxing temperature until the TLC analysis indicated the completion of the reaction (see Table 1 for reaction times).

Method B: A solution of **3a** or **3b** (128 mg, 0.33 mmol) and the aldehyde (0.50 mmol) in dry CH<sub>2</sub>Cl<sub>2</sub> (10 mL) in the presence of powdered 4 Å MS was stirred at room temperature until TLC analysis indicated the completion of the reaction (see Table 1 for reaction times). The reaction mixture was filtered through a pad of Celite with washing by diethyl ether. The combined organic solution was evaporated under reduced pressure. The residue was purified by flash column chromatography (silica gel, 14% EtOAc in hexane) to give the oxazolidines. The yields and specific rotation data are summarized in Table 1.

# 3.1.1. (2S,4S)-4-(Indol-3-yl-methyl)-2,3-dimethyl-5,5-di(p-tolyl)-1,3-oxazolidine 1a

Pale yellow foam; IR (KBr) 3420 (br), 2920, 1455, 1350, 820, 750 cm $^{-1}$ ;  $^{1}$ H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.96 (s, 1H), 7.60 (d, J=7.8 Hz, 1H), 7.35 (d, J=8.1 Hz, 1H), 7.24–7.05 (m, 10H), 6.58 (s, 1H), 4.00 (q, J=5.1 Hz, 1H), 3.68 (t, J=6.3 Hz, 1H), 2.76 (d, J=6.3 Hz, 2H), 2.35 (s, 3H), 2.33 (s, 3H), 2.22 (s, 3H), 1.54 (d, J=5.1 Hz, 3H);  $^{13}$ C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  142.9, 140.5, 136.7, 136.5, 136.2, 128.6, 128.4, 128.1, 127.5, 126.9, 122.7, 121.8, 119.2, 119.0, 113.1, 111.1, 91.5, 88.1, 73.8, 37.8, 29.0, 21.0, 20.9, 19.0; MS (CI $^{+}$ ) m/z (relative intensity) 411 (M $^{+}$ +1, 100); HRMS (FAB $^{+}$ ) calcd for C<sub>28</sub>H<sub>31</sub>N<sub>2</sub>O (M $^{+}$ +1): 411.2436. Found: 411.2484. Anal. calcd for C<sub>28</sub>H<sub>30</sub>N<sub>2</sub>O: C, 81.91; H, 7.37; N, 6.82. Found: C, 81.79; H, 7.55; N, 6.50.

# 3.1.2. (2S,4S)-2-Ethyl-4-(indol-3-yl-methyl)-3-methyl-5,5-di(p-tolyl)-1,3-oxazolidine 1b

Pale yellow foam; IR (KBr) 3395, 2920, 1450, 1340, 1230, 1180, 1010, 810, 740 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.90 (s, 1H), 7.60 (d, J=7.9 Hz, 1H), 7.36 (d, J=8.1 Hz, 1H), 7.26–7.06 (m, 10H), 6.61 (d, J=2.3 Hz, 1H), 3.85 (dd, J=6.4, 2.9 Hz, 1H), 3.71 (t, J=6.4 Hz, 1H), 2.69 (dd, J=7.0, 2.3 Hz, 2H), 2.35 (s, 3H), 2.33 (s, 3H), 2.17 (s, 3H), 1.90–1.61 (m, 2H), 1.17 (t, J=7.4 Hz, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  142.9, 140.6, 136.5, 136.4, 136.2, 128.7, 128.2, 128.0, 127.6, 126.9, 122.7, 121.8, 119.2, 119.0, 113.5, 111.0, 95.9, 87.8, 73.4, 38.7, 29.4, 26.5, 21.0, 20.9, 9.1; HRMS (FAB<sup>+</sup>) calcd for C<sub>29</sub>H<sub>33</sub>N<sub>2</sub>O (M<sup>+</sup>+1): 425.2593. Found: 425.2644. Anal. calcd for C<sub>29</sub>H<sub>32</sub>N<sub>2</sub>O: C, 82.04; H, 7.60; N, 6.60. Found: C, 82.13; H, 7.65; N, 6.44.

# 3.1.3. (2S,4S)-4-(Indol-3-yl-methyl)-3-methyl-2-propyl-5,5-di(p-tolyl)-1,3-oxazolidine 1c

Pale yellow foam; IR (KBr) 3400, 2920, 1450, 1350, 1180, 1020, 810, 740 cm<sup>-1</sup>;  $^{1}$ H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.91 (s, 1H), 7.58 (d, J=7.5 Hz, 1H), 7.35 (d, J=7.8 Hz, 1H), 7.26–7.04 (m, 10H), 6.59 (s, 1H), 3.87 (t, J=4.5 Hz, 1H), 3.67 (t, J=6.3 Hz, 1H), 2.69 (d, J=6.0 Hz, 2H), 2.33 (s, 3H), 2.31 (s, 3H), 2.17 (s, 3H), 1.83–1.56 (m, 4H), 1.03 (t, J=7.5 Hz, 3H);  $^{13}$ C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  142.8, 140.5, 136.5, 136.4, 136.2, 128.7, 128.2, 128.1, 127.4, 126.9, 122.8, 121.8, 119.2, 119.0, 113.6, 111.0, 95.0, 87.8, 73.5, 38.6, 35.7, 29.3, 21.0, 20.9, 18.3, 14.4; MS (CI<sup>+</sup>) m/z (relative intensity) 439 (M<sup>+</sup>+1, 100); HRMS (FAB<sup>+</sup>) calcd for C<sub>30</sub>H<sub>35</sub>N<sub>2</sub>O (M<sup>+</sup>+1): 439.2749. Found: 439.2720. Anal. calcd for C<sub>30</sub>H<sub>34</sub>N<sub>2</sub>O: C, 82.15; H, 7.81; N, 6.39. Found: C, 82.03; H, 7.99; N, 6.22.

# 3.1.4. (2S,4S)-2-Butyl-4-(indol-3-yl-methyl)-3-methyl-5,5-di(p-tolyl)-1,3-oxazolidine 1d

Pale yellow foam; IR (KBr) 3400, 2920, 1450, 1350, 1180, 1020, 820, 745 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.93 (s, 1H), 7.59 (d, J=7.8 Hz, 1H), 7.35 (d, J=8.1 Hz, 1H), 7.26–7.05 (m, 10H), 6.58 (s, 1H), 3.86 (dd, J=6.0, 3.0 Hz, 1H), 3.68 (t, J=6.3 Hz, 1H), 2.70 (d, J=6.0 Hz, 2H), 2.34 (s, 3H), 2.32 (s, 3H), 2.18 (s, 3H), 1.84–1.28 (m, 6H), 0.98 (t, J=7.2 Hz, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  142.9, 140.5, 136.5, 136.4, 136.2, 128.7, 128.2, 128.1, 127.6, 126.8, 122.8, 121.8, 119.1, 118.9, 113.3, 111.0, 95.1, 87.8, 73.5, 38.6, 33.3, 29.3, 27.2, 23.0, 21.0, 20.9, 14.1; MS (CI<sup>+</sup>) m/z (relative intensity) 453 (M<sup>+</sup>+1, 100); HRMS (FAB<sup>+</sup>) calcd for C<sub>31</sub>H<sub>37</sub>N<sub>2</sub>O (M<sup>+</sup>+1): 453.2906. Found: 453.2924. Anal. calcd for C<sub>31</sub>H<sub>36</sub>N<sub>2</sub>O: C, 82.26; H, 8.02; N, 6.19. Found: C, 82.15; H, 8.06; N, 6.04.

# 3.1.5. (2S,4S)-4-(Indol-3-yl-methyl)-3-methyl-2-pentyl-5,5-di(p-tolyl)-1,3-oxazolidine 1e

Pale yellow foam; IR (KBr) 3400, 2910, 2840, 1440, 1340, 1320, 1180, 1005, 805, 730 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.00 (s, 1H), 7.60 (d, J=7.5 Hz, 1H), 7.33 (d, J=7.8 Hz, 1H), 7.30–7.07 (m, 10H), 6.58 (s, 1H), 3.89 (dd, J=6.0, 3.0 Hz, 1H), 3.71 (t, J=6.2 Hz, 1H), 2.73 (d, J=6.0 Hz, 2H), 2.34 (s, 3H), 2.33 (s, 3H), 2.19 (s, 3H), 2.00–1.25 (m, 8H), 0.96 (t, J=7.0 Hz, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 143.0, 140.6, 136.5, 136.4, 136.3, 128.7, 128.2, 128.0, 127.6, 126.7, 122.8, 121.7, 119.1, 118.9, 113.4,

111.0, 95.2, 87.9, 73.4, 38.6, 33.7, 32.1, 29.7, 29.4, 24.6, 22.7, 20.9, 14.0; HRMS (FAB<sup>+</sup>) calcd for  $C_{32}H_{39}N_2O$  (M<sup>+</sup>+1): 467.3062. Found: 467.3101.

- 3.1.6. (2S,4S)-4-(Indol-3-yl-methyl)-2-isopropyl-3-methyl-5,5-di(p-tolyl)-1,3-oxazolidine If Pale yellow foam; IR (KBr) 3395, 2940, 1445, 1340, 1180, 810, 740 cm $^{-1}$ ;  $^{1}$ H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.86 (s, 1H), 7.58 (d, J=7.5 Hz, 1H), 7.36–7.08 (m, 11H), 6.73 (d, J=2.1 Hz, 1H), 3.80 (dd, J=8.1, 5.4 Hz, 1H), 3.75 (d, J=3.6 Hz, 1H), 2.58 (dd, J=14.4, 8.4 Hz, 1H), 2.52 (dd, J=14.1, 5.4 Hz, 1H), 2.32 (s, 6H), 2.04 (s, 3H), 1.95 (d of septet, J=3.6, 6.9 Hz, 1H), 1.20 (d, J=6.9 Hz, 3H), 1.12 (d, J=6.9 Hz, 3H);  $^{13}$ C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  142.8, 140.6, 136.4, 136.1, 136.0, 128.8, 128.3, 127.7, 127.2, 126.7, 122.7, 121.6, 119.0, 118.8, 113.9, 111.0, 99.0, 87.6, 72.4, 40.6, 31.7, 29.8, 21.0, 20.9, 18.7, 16.5; MS (CI $^{+}$ ) m/z (relative intensity) 439 (M $^{+}$ +1, 100); HRMS (FAB $^{+}$ ) calcd for C<sub>30</sub>H<sub>35</sub>N<sub>2</sub>O (M $^{+}$ +1): 439.2749. Found: 439.2721. Anal. calcd for C<sub>30</sub>H<sub>34</sub>N<sub>2</sub>O: C, 82.15; H, 7.81; N, 6.39. Found: C, 82.11; H, 7.93; N, 6.18.
- 3.1.7. (2S,4S)-4-(Indol-3-yl-methyl)-3-methyl-2-(2-methylpropyl)-5,5-di(p-tolyl)-1,3-oxazolidine 1g Pale yellow foam; IR (KBr) 3400, 2940, 1450, 1340, 1180, 1010, 810, 740 cm $^{-1}$ ;  $^{1}$ H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.90 (s, 1H), 7.60 (d, J=7.8 Hz, 1H), 7.37 (d, J=8.0 Hz, 1H), 7.24–7.06 (m, 10H), 6.60 (d, J=1.8 Hz, 1H), 3.91 (dd, J=8.3, 2.4 Hz, 1H), 3.67 (t, J=6.5 Hz, 1H), 2.70 (d, J=6.5 Hz, 2H), 2.34 (s, 3H), 2.32 (s, 3H), 2.18 (s, 3H), 2.10–1.95 (m, 1H), 1.85–1.76 (m, 1H), 1.70–1.61 (m, 1H), 1.07 (d, J=6.7 Hz, 3H), 0.99 (d, J=6.6 Hz, 3H);  $^{13}$ C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  143.1, 140.6, 136.5, 136.4, 136.2, 128.7, 128.2, 128.1, 127.6, 126.8, 122.7, 121.8, 119.2, 119.0, 113.4, 111.0, 93.9, 88.0, 73.4, 42.9, 38.5, 29.4, 25.2, 23.8, 22.6, 21.0, 20.9; HRMS (FAB $^{+}$ ) calcd for C<sub>31</sub>H<sub>37</sub>N<sub>2</sub>O (M $^{+}$ +1): 453.2906. Found: 453.2978. Anal. calcd for C<sub>31</sub>H<sub>36</sub>N<sub>2</sub>O: C, 82.26; H, 8.02; N, 6.19. Found: C, 82.25; H, 8.06; N, 6.06.
- 3.1.8. (2S,4S)-4-(Indol-3-yl-methyl)-3-methyl-2-(2,2-dimethylpropyl)-5,5-di(p-tolyl)-1,3-oxazolidine  $\it Ih$  Pale yellow foam; IR (KBr) 3430, 2960, 1420, 1320, 840, 700 cm $^{-1}$ ;  $^{1}H$  NMR (300 MHz, CDCl $_3$ )  $\delta$  7.91 (s, 1H), 7.61 (d,  $\it J$ =7.7 Hz, 1H), 7.37 (d,  $\it J$ =8.0 Hz, 1H), 7.26–7.06 (m, 10H), 6.62 (d,  $\it J$ =2.2 Hz, 1H), 3.92 (dd,  $\it J$ =8.0, 1.0 Hz, 1H), 3.65 (t,  $\it J$ =6.5 Hz, 1H), 2.68 (d,  $\it J$ =6.4 Hz, 2H), 2.34 (s, 3H), 2.33 (s, 3H), 2.17 (s, 3H), 1.85 (dd,  $\it J$ =14.3, 8.2 Hz, 1H), 1.72 (d,  $\it J$ =14.3 Hz, 1H), 1.06 (s, 9H);  $\it I$ 3C NMR (75 MHz, CDCl $_3$ )  $\delta$  143.2, 140.8, 136.4, 136.3, 136.0, 128.7, 128.2, 128.1, 127.7, 126.9, 122.7, 121.8, 119.2, 119.0, 113.4, 111.0, 93.1, 88.5, 72.7, 47.3, 38.3, 30.2, 29.8, 29.5, 21.0, 20.9; HRMS (FAB $^+$ ) calcd for C $_3$ 2H $_3$ 9N $_2$ O (M $^+$ +1): 467.3062. Found: 467.3129. Anal. calcd for C $_3$ 2H $_3$ 8N $_2$ O: C, 82.36; H, 8.21; N, 6.00. Found: C, 82.20; H, 8.28; N, 5.90.
- 3.1.9. (2S,4S)-4-(Indol-3-yl-methyl)-3-methyl-2-(2-phenylethyl)-5,5-di(p-tolyl)-1,3-oxazolidine Ii
  Pale yellow foam; IR (KBr) 3400, 2940, 1455, 1355, 830, 745 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ
  7.89 (s, 1H), 7.60 (d, *J*=7.8 Hz, 1H), 7.37–7.07 (m, 16H), 6.62 (d, *J*=1.8 Hz, 1H), 3.95 (dd, *J*=6.3, 2.7 Hz, 1H), 3.73 (t, *J*=6.3 Hz, 1H), 3.17–3.04 (m, 1H), 2.96–2.82 (m, 1H), 2.69 (d, *J*=6.3 Hz, 2H), 2.35 (s, 3H), 2.33 (s, 3H), 2.15 (s, 3H), 2.23–2.04 (m, 2H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 142.8, 142.5, 140.5, 136.6, 136.5, 136.2, 128.7, 128.5, 128.3, 128.2, 128.0, 127.5, 126.9, 125.7, 122.7, 121.8, 119.2, 118.9, 113.4, 111.0, 94.3, 88.1, 73.2, 38.7, 35.3, 31.0, 29.5, 21.0, 20.9; MS (CI<sup>+</sup>) *m/z* (relative intensity) 501 (M<sup>+</sup>+1, 100); HRMS (FAB<sup>+</sup>) calcd for C<sub>35</sub>H<sub>37</sub>N<sub>2</sub>O (M<sup>+</sup>+1): 501.2906. Found: 501.2976. Anal. calcd for C<sub>35</sub>H<sub>36</sub>N<sub>2</sub>O: C, 83.96; H, 7.25; N, 5.60. Found: C, 83.77; H, 7.39; N, 5.43.

3.1.10. (2S,4S)-4-(Indol-3-yl-methyl)-3-methyl-2-phenyl-5,5-di(p-tolyl)-1,3-oxazolidine Ij

Pale yellow foam; IR (KBr) 3410, 2940, 2870, 1670, 1460, 1355, 830, 745 cm<sup>-1</sup>;  $^{1}$ H NMR (300 MHz, CDCl<sub>3</sub>) δ 7.90 (s, 1H), 7.71 (d, J=6.3 Hz, 2H), 7.58 (d, J=7.8 Hz, 1H), 7.49–7.07 (m, 14H), 6.59 (d, J=1.5 Hz, 1H), 4.72 (s, 1H), 3.87 (t, J=6.3 Hz, 1H), 2.81 (ABX, J=11.1, 6.9 Hz, 2H), 2.34 (s, 3H), 2.33 (s, 3H), 2.12 (s, 3H);  $^{13}$ C NMR (75 MHz, CDCl<sub>3</sub>) δ 142.6, 140.4, 139.0, 136.6, 136.5, 136.2, 129.0, 128.8, 128.4, 128.4, 128.2, 128.2, 127.5, 127.1, 122.8, 121.8, 119.2, 118.9, 113.2, 111.1, 96.4, 88.9, 72.9, 37.8, 29.4, 21.0, 20.9; MS (CI<sup>+</sup>) m/z (relative intensity) 473 (M<sup>+</sup>+1, 50), 211 (100); HRMS (FAB<sup>+</sup>) calcd for C<sub>33</sub>H<sub>33</sub>N<sub>2</sub>O (M<sup>+</sup>+1): 473.2593. Found: 473.2611. Anal. calcd for C<sub>33</sub>H<sub>32</sub>N<sub>2</sub>O: C, 83.86; H, 6.82; N, 5.93. Found: C, 83.79; H, 6.98; N, 5.71.

# 3.1.11. (2S,4S)-4-(Indol-3-yl-methyl)-2,3-dimethyl-5,5-di(0-tolyl)-1,3-oxazolidine 2a

Pale yellow foam; IR (KBr) 3250, 2925, 1457, 1341, 1234, 1140, 1071, 750, 735 cm $^{-1}$ ;  $^{1}$ H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.89 (s, 1H), 7.60 (d, J=7.7 Hz, 1H), 7.49 (d, J=6.3 Hz, 1H), 7.32 (d, J=8.0 Hz, 1H), 7.18–6.99 (m, 9H), 6.80 (s, 1H), 4.28 (d, J=8.7 Hz, 1H), 3.76 (q, J=5.0 Hz, 1H), 2.36 (dd, J=11.3, 8.7 Hz, 1H), 2.26 (d, J=11.3 Hz, 1H), 2.08 (s, 6H), 2.00 (s, 3H), 1.37 (d, J=5.0 Hz, 3H);  $^{13}$ C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  140.8, 139.1, 136.4, 133.2, 131.6, 128.5, 127.6, 127.2, 125.1, 124.0, 121.6, 119.2, 118.7, 111.1, 90.3, 89.5, 67.6, 38.4, 29.7, 21.8, 14.1; HRMS (FAB $^{+}$ ) calcd for C<sub>28</sub>H<sub>31</sub>N<sub>2</sub>O (M $^{+}$ +1): 411.2436. Found: 411.2347.

# 3.1.12. (2S,4S)-2-Ethyl-4-(indol-3-yl-methyl)-3-methyl-5,5-di(0-tolyl)-1,3-oxazolidine 2b

Pale yellow foam; IR (KBr) 3424, 2967, 2930, 1484, 1456, 1353, 1230, 1068, 1014, 751, 742 cm<sup>-1</sup>;  $^{1}$ H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.91 (s, 1H), 7.59 (d, J=7.7 Hz, 1H), 7.53 (d, J=7.0 Hz, 1H), 7.31 (d, J=8.0 Hz, 1H), 7.21–7.00 (m, 9H), 6.85 (s, 1H), 4.34 (d, J=8.7 Hz, 1H), 3.65 (dd, J=5.8, 2.8 Hz, 1H), 2.36 dd, J=12.2, 8.7 Hz, 1H), 2.21 (d, J=12.2 Hz, 1H), 2.08 (s, 6H), 2.04 (s, 3H), 1.78–1.60 (m, 2H), 1.08 (t, J=7.4 Hz, 3H);  $^{13}$ C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  141.0, 139.1, 136.3, 134.8, 133.1, 131.4, 129.0, 128.3, 127.9, 127.4, 126.9, 124.9, 123.7, 123.0, 121.5, 119.0, 118.8, 114.2, 111.0, 94.4, 89.8, 66.8, 38.9, 31.2, 27.2, 22.3, 21.7, 8.7; HRMS (FAB<sup>+</sup>) calcd for  $C_{29}H_{33}N_{2}O$  (M<sup>+</sup>+1): 425.2593. Found: 425.2681.

# 3.1.13. (2S,4S)-4-(Indol-3-yl-methyl)-3-methyl-2-propyl-5,5-di(o-tolyl)-1,3-oxazolidine 2c

Pale yellow foam; IR (KBr) 3421, 3259, 2959, 1457, 1356, 1233, 1137, 1016, 754, 739 cm<sup>-1</sup>;  $^{1}$ H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.94 (s, 1H), 7.60 (d, J=7.7 Hz, 1H), 7.53 (d, J=6.9 Hz, 1H), 7.33 (d, J=8.0 Hz, 1H), 7.22–7.00 (m, 9H), 6.83 (s, 1H), 4.32 (d, J=8.6 Hz, 1H), 3.66 (dd, J=5.6, 2.7 Hz, 1H), 2.34 (dd, J=12.1, 8.6 Hz, 1H), 2.22 (d, J=12.1 Hz, 1H), 2.09 (s, 6H), 2.05 (s, 3H), 2.03–1.46 (m, 4H), 1.00 (t, J=7.1 Hz, 3H);  $^{13}$ C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  141.0, 139.1, 136.3, 134.8, 133.0, 131.4, 129.0, 128.4, 127.9, 127.4, 126.9, 124.9, 123.7, 123.0, 121.5, 119.0, 118.8, 114.2, 111.0, 93.6, 89.7, 66.7, 38.9, 36.9, 31.2, 22.3, 21.6, 18.0, 14.4; HRMS (FAB<sup>+</sup>) calcd for  $C_{30}H_{35}N_2O$  (M<sup>+</sup>+1): 439.2749. Found: 439.2809.

# 3.1.14. (2S,4S)-2-Butyl-4-(indol-3-yl-methyl)-3-methyl-5,5-di(0-tolyl)-1,3-oxazolidine 2d

Pale yellow foam; IR (KBr) 3226, 2954, 2928, 1458, 1353, 1234, 1131, 1017, 754, 739 cm<sup>-1</sup>;  $^{1}$ H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.88 (s, 1H), 7.59 (d, J=7.8 Hz, 1H), 7.52 (d, J=6.7 Hz, 1H), 7.33 (d, J=8.0 Hz, 1H), 7.21–7.00 (m, 9H), 6.83 (s, 1H), 4.31 (d, J=8.6 Hz, 1H), 3.65 (dd, J=6.0, 2.9 Hz, 1H), 2.34 (dd, J=11.8, 8.6 Hz, 1H), 2.20 (d, J=11.8 Hz, 1H), 2.16 (s, 6H), 2.04 (s, 3H), 2.03–1.32 (m, 6H), 0.95 (t, J=7.1 Hz, 3H);  $^{13}$ C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  140.3, 139.1, 136.3, 133.3, 131.6, 128.4, 127.7, 127.2, 125.1, 123.9, 121.6, 119.2, 118.4, 111.2, 94.0, 90.1, 67.5, 38.9, 33.5, 27.1, 22.8, 22.5, 21.6, 14.1; HRMS (FAB+) calcd for  $C_{31}H_{37}N_{2}O$  (M<sup>+</sup>+1): 453.2906. Found: 453.2885.

- 3.1.15. (2S,4S)-4-(Indol-3-yl-methyl)-3-methyl-2-pentyl-5,5-di(o-tolyl)-1,3-oxazolidine **2e** Pale yellow foam; IR (KBr) 3221, 2953, 2928, 1458, 1353, 1129, 1017, 754, 734 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.88 (s, 1H), 7.59 (d, J=7.8 Hz, 1H), 7.52 (d, J=6.7 Hz, 1H), 7.33 (d, J=8.0 Hz, 1H), 7.21–7.00 (m, 9H), 6.83 (s, 1H), 4.31 (d, J=8.6 Hz, 1H), 3.65 (dd, J=6.0, 2.9 Hz, 1H), 2.34 (dd, J=11.8, 8.6 Hz, 1H), 2.20 (d, J=11.8 Hz, 1H), 2.16 (s, 6H), 2.04 (s, 3H), 2.03–1.32 (m, 8H), 0.90 (t, J=7.1 Hz, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  141.0, 139.1, 136.3, 135.0, 133.1, 131.4, 129.0, 128.4, 127.7, 127.4, 126.7, 124.9, 123.7, 122.9, 121.5, 119.0, 118.9, 114.3, 111.0, 93.8, 89.8, 66.9, 38.9, 34.4, 31.2, 24.4, 23.0, 22.3, 21.7, 14.2; HRMS (FAB<sup>+</sup>) calcd for C<sub>32</sub>H<sub>39</sub>N<sub>2</sub>O (M<sup>+</sup>+1): 467.3062. Found: 467.3066.
- 3.1.16. (2S,4S)-4-(Indol-3-yl-methyl)-2-isopropyl-3-methyl-5,5-di(o-tolyl)-1,3-oxazolidine **2f** Pale yellow foam; IR (KBr) 3417, 3392, 2960, 2927, 1456, 1357, 1230, 1054, 1023, 746 cm $^{-1}$ ;  $^{1}$ H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.93 (s, 1H), 7.57 (d, J=7.5 Hz, 2H), 7.34 (d, J=8.0 Hz, 1H), 7.30–7.00 (m, 9H), 6.95 (s, 1H), 4.42 (br s, 1H), 3.51 (d, J=3.4 Hz, 1H), 2.44 (br s, 1H), 2.19 (d, J=13.2 Hz, 1H), 2.13 (s, 3H), 2.06 (s, 6H), 2.00–1.80 (m, 1H), 1.16 (d, J=6.8 Hz, 3H), 1.04 (d, J=6.9 Hz, 3H);  $^{13}$ C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  140.9, 139.2, 136.3, 134.6, 133.2, 131.4, 129.1, 128.2, 127.8, 127.5, 127.0, 125.0, 123.7, 123.2, 121.5, 119.1, 118.6, 114.1, 111.1, 98.0, 89.4, 66.8, 40.2, 31.7, 30.8, 29.7, 22.3, 21.5, 18.7; HRMS (FAB+) calcd for C<sub>30</sub>H<sub>35</sub>N<sub>2</sub>O (M++1): 439.2749. Found: 439.2685.
- 3.1.17. (2S,4S)-4-(Indol-3-yl-methyl)-3-methyl-2-(2-methylpropyl)-5,5-di(o-tolyl)-1,3-oxazolidine **2g** Pale yellow foam; IR (KBr) 3428, 2954, 2928, 1456, 1355, 1232, 1014, 745 cm $^{-1}$ ;  $^{1}$ H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.92 (s, 1H), 7.59 (d, J=7.7 Hz, 1H), 7.51 (d, J=7.2 Hz, 1H), 7.32 (d, J=7.9 Hz, 1H), 7.25–7.00 (m, 9H), 6.87 (br s, 1H), 4.44 (br s, 1H), 3.70 (d, J=7.3 Hz, 1H), 2.40 (br s, 1H), 2.25 (d, J=13.4 Hz, 1H), 2.11 (s, 3H), 2.04 (s, 6H), 2.00–1.50 (m, 3H), 1.01 (d, J=6.7 Hz, 3H), 0.86 (d, J=6.5 Hz, 3H);  $^{13}$ C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  140.5, 139.0, 136.3, 133.2, 131.5, 128.8, 128.5, 127.5, 127.0, 125.0, 123.8, 123.3, 121.6, 119.1, 118.7, 111.0, 92.9, 90.0, 67.0, 43.9, 38.8, 29.7, 25.1, 23.7, 22.7, 22.4, 21.6; HRMS (FAB $^{+}$ ) calcd for C<sub>31</sub>H<sub>37</sub>N<sub>2</sub>O (M $^{+}$ +1): 453.2906. Found: 453.2948.

# 3.2. A typical procedure for the catalytic addition of $Et_2Zn$ to bezaldehyde

To a solution of the chiral oxazolidine 1 or 2 (0.18 mmol) in dry PhMe (8 mL) under a nitrogen atmosphere cooled in an ice-water bath (ca. 0°C) was added a solution of Et<sub>2</sub>Zn (4 mL, 1 M in hexanes) via a syringe. After stirring for 10 min, freshly distilled benzaldehyde (0.20 mL, 1.80 mmol) was added into the mixture via another syringe. The resultant mixture was allowed to warm up to room temperature and stirring was continued for 94–100 h. The reaction mixture was cooled in an ice-water bath and quenched with 5% HCl aqueous solution. The mixture was extracted with diethyl ether (3×20 mL), washed with brine, dried over anhydrous MgSO<sub>4</sub>, and condensed under reduced pressure. The residue was purified by flash column chromatography (silica gel, 10% EtOAc in hexane) to give chiral 1-phenyl-1-propanol (4). The yields and enantiomeric excess data are summarized in Table 2.

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